

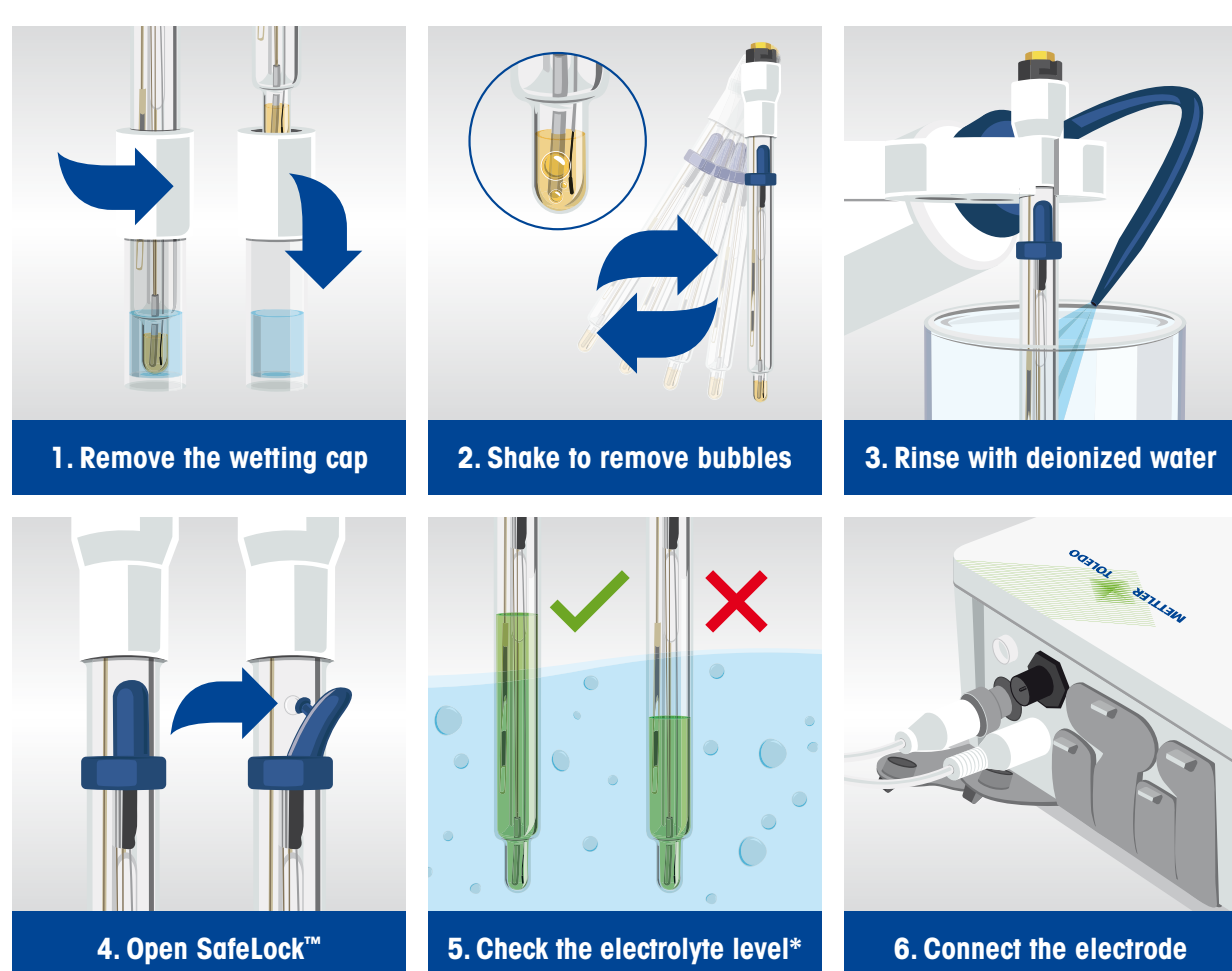


# pH Electrode Maintenance Guide

## Electrode Preparation

## Electrode Calibration

Before use



[www.mt.com/phlab-electrodepreparation](http://www.mt.com/phlab-electrodepreparation)

\* Artificially colored green for illustration purpose

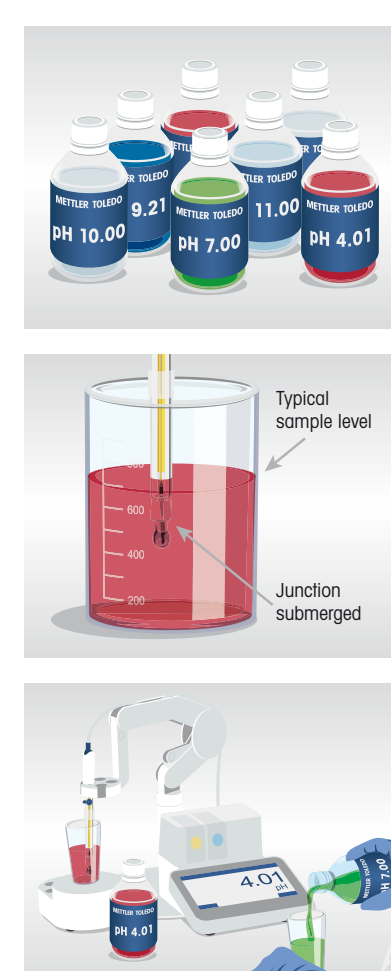
1. Take a minimum of two pH buffers bracketing the sample pH.
2. Use a fresh amount of aliquot of non-expired pH buffer for every calibration point.
3. Pour enough to submerge the sensor junction.
4. Stir the buffers (if required).
5. Start the calibration and take the temperature of buffers into account.
6. Rinse electrode thoroughly with deionized water and blot dry with tissue.
7. Repeat steps 2–6 for next buffer.
8. Evaluate as shown in table below.

Slope/Offset	± 0–20 mV	± 20–35 mV	> 35 mV
95–105%	✓	✓	✓
90–95%	✓	✓	✓
85–90%	✓	✓	✓
< 85% or > 105%	✗	✗	✗

- ✓ Electrode in good condition
- ✓ Electrode requires cleaning soon\*
- ✓ Cleaning and regeneration required\*
- ✗ Electrode worn out and needs replacement



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\* Refer to the Electrode Troubleshooting section below

## Good Measuring Practices (pH/Titration)

During use

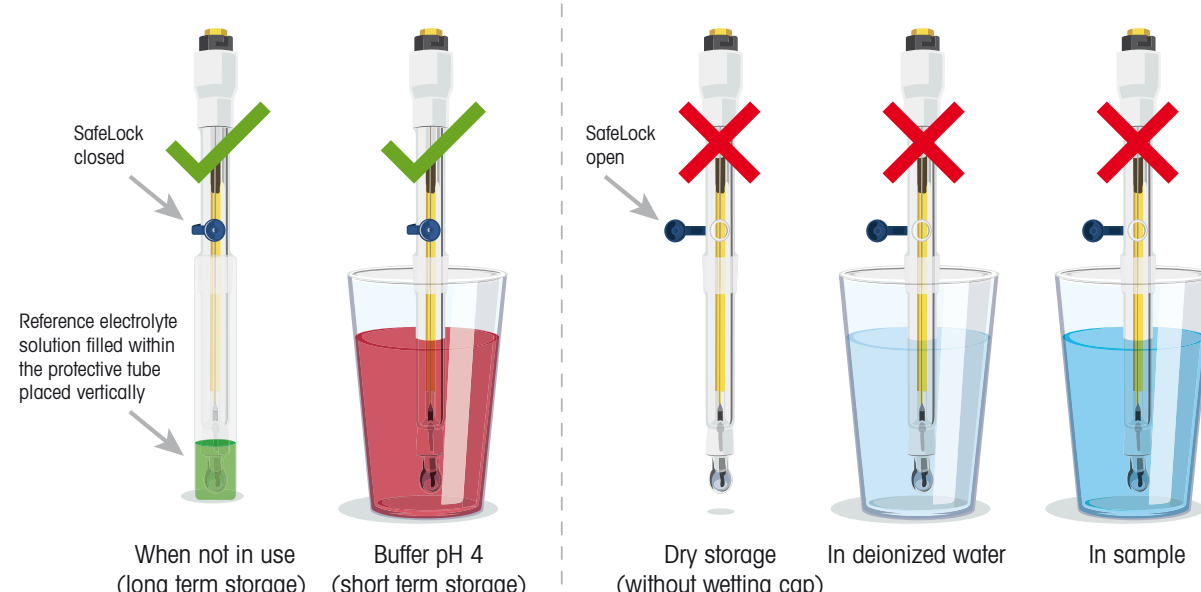
1. Take suitable electrode according to the sample to be measured.
2. Rinse the electrode with deionized water and blot dry with tissue.
3. Ensure that the electrode junction is immersed in sample/buffer.
4. Ensure that the reference electrolyte level is above the sample solution level.
5. Calibrate the sensor before use.
6. Perform kinetic electrode test if required (for titration only).
7. Maintain uniform stirring and temperature conditions for calibration and sample measurement (pH/Titration).
8. Rinse the electrode thoroughly between consecutive measurements.



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## Electrode Storage

After use



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- Long term** In reference electrolyte solution (specific for each electrode).
- Short term** Buffer pH 4/7.
- Never** Never store the electrode dry or in distilled water as this will affect the pH-sensitive glass membrane and thus shorten the lifetime of the electrode.

## Electrode Troubleshooting

### 1. Symptoms, causes and regeneration procedure for pH sensitive glass membrane

Inspection	Cause	Ageing of glass	Scratches on membrane	Broken membrane or shaft	Gel layer destroyed or dehydrated	Dry storage of electrode	Calcium on glass membrane (whitish film)	Oil, fat or tar residues (visible?)	Deposits of unknown substances (visible?)
<b>Environment</b> <b>Is instrument ok?</b> • Cross-check with another instrument  <b>Is electrode cable ok?</b> • Cross-check with another cable	Reduced slope (80–90%)	●	●	—	—	—	●	●	●
	Very small slope (<80%)	●	—	●	●	●	—	—	—
	Slow response	●	●	—	●	●	●	●	●
	Fluctuating reading	●	●	●	●	●	●	●	●
	Zero point shift	●	—	—	—	●	●	●	●
	Reason	High temperature, age of electrode	Abrasion, solid particles, incorrect cleaning	Mechanical or temperature shock	Ion deficient media, non-aqueous applications	Wrong storage	Measurement media	Measurement media	Measurement media, no cleaning
<b>Electrode</b>  <b>Presence of an air bubble?</b> • Refer image 2 in electrode preparation  <b>Reference electrolyte level?</b> • Refer image 5 in electrode preparation  <b>Glass membrane activation?</b> • Refer table 1 in this section  <b>Clogged junction?</b> • Refer table 2 in this section	Restoration procedures	Regeneration*	Replace electrode	Replace electrode	Rehydration with electrolyte	Rehydration with electrolyte	Dip the glass in acetic acid / dilute hydrochloric acid until deposits are dissolved. Follow up with regeneration	Clean with degreasing agent, then with deionized water. Condition the electrode in pH buffer 4	Clean with degreasing agent, then with deionized water. Condition the electrode in pH buffer 4
		● Possible ● Probable ● Highly probable	* Immerse the electrode glass membrane in regeneration solution for not more than 2 minutes						

### 2. Clogged junction: Causes and cleaning procedures

Type of contamination	Cleaning agent	Reaction time	Remarks
Silver sulfide	Thiourea	5–60 minutes	Leave until coloration disappears
All possible types of contamination	First recommendation for removing unknown substances HCl 0.1 M	Approximately 12 hours	Can also be used for internal cleaning
	Second recommendation for removing unknown substances Chromic-sulfuric acid mixture	Approximately 30 minutes	Also cleans deposits very well. Electrode must be regenerated after this
Proteins	Pepsin/HCl cleaning solution	>1 hour	Can also be used for internal cleaning
	NaOH 2%	Approximately 20 minutes	Rinse the electrode with deionized water followed by conditioning in pH buffer 4
Lipophilic substances	Ethanol, acetone	Approximately 30 minutes	Highly suitable for edible oils. Always recondition the pH electrode in buffer pH 4 or 0.1M HCl
Calcium, scale	Acetic acid	Approximately 30 minutes	Electrode must be regenerated after this procedure
Soaps, tensides	Hot water (80 °C)	Approximately 12 hours	Thorough washing with deionized water followed by conditioning in pH buffer 4



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